

Introduction

The ICH S7B and E14 guidelines, which focus on hERG channel block and QT prolongation, are highly sensitive but not very specific for early prediction of proarrhythmic risks. Opposed to those guidelines, the Comprehensive in vitro Proarrhythmia Assay (CiPA) was proposed to determine whether new drug candidates are proarrhythmic by integrating the drug effects on multiple human cardiac ion channels into a human cardiomyocyte model. This approach has a potential to improve sensitivity as well as specificity for proarrhythmic risks. Through the validation process of this model, the CiPA initiative has concluded that three ion channels, hERG, hNav1.5 (late current), and hCav1.2, have significant impacts on the action potential duration of cardiomyocytes. Many researchers worldwide have attempted for verification of the CiPA-recommended protocols of 2018, to our knowledge, yet few ion channel data are publicly available. The purpose of this research is to collect ion channel data with sufficient accuracy, faithfully based on the CiPA protocols, to fulfill precise proarrhythmic prediction. We performed block potency measurement with several CiPA training/validation drugs on our manual patch-clamp platform in HEK293/CHO cells expressing hERG, hNav1.5, and hCav1.2. Of the CiPA drugs, the following was selected: bepridil, dofetilide, chlorpromazine, cisapride, astemizole, ranolazine and verapamil; two or three drugs each from the high, intermediate, and low TdP risk categories. IC50 values / Hill coefficients were calculated from the results. We would like to present and discuss differences in the parameter sets from the CiPA datasets including high-throughput platform. In silico prediction will be performed with the data set obtained in the research.

Materials & Methods

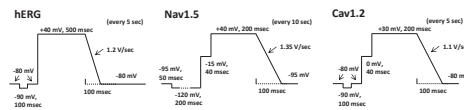
CiPA compounds used:

TdP risk	High	Intermediate	Low
	Bepridil Dofetilide	Astemizole Chlorpromazine Cisapride	Ranolazine Verapamil

Cell lines: hERG-HEK293, Nav1.5-HEK293, Cav1.2-CHO.

Patch-clamp recording: Voltage protocols, temperature, and ionic compositions of the solutions were fully identical to what the CiPA initiative announced. In a subset of assays, sucrose was added to the extracellular solutions for optimizing the osmotic balance.

Voltage protocols

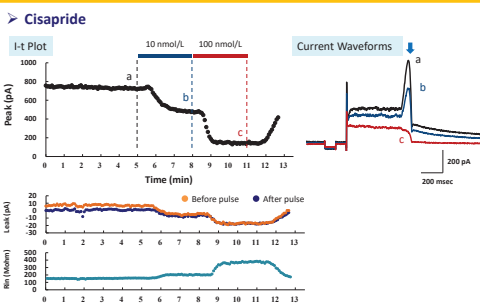


hERG dynamic model protocols



Results

hERG Assay with IC50 Only Protocol



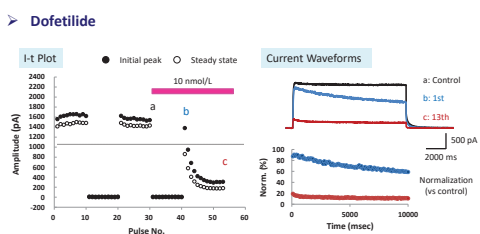
Note: Many cells showed relatively lower input resistance and outward leak currents in spite of the holding potential of -80 mV. When the cells were exposed to intensively inhibiting compounds, the input resistance was gradually increased along with decline of leak current to less than 0 pA.

Table 1. Effects of CiPA Compounds on hERG Channel (IC50 only protocol)

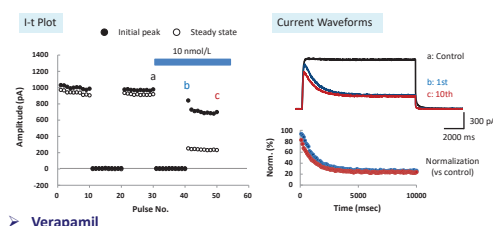
TdP risk	Test concentration (μmol/L)	n	IC50	Hill	CiPA data set			
					Manual ¹	Hill	HTS ²	
Bepridil	3, 10, 30, 100	4	98.38	0.97	100	0.98	264	1.4
Dofetilide ^a	3, 10, 30, 100	4	21.67	1.29	1.47	0.62	12.4	2.4
Astemizole ^a	0.1, 0.3, 1, 3	4	1.00	0.99	9.99	0.54	23.3	5.4
Chlorpromazine ^a	100, 300, 1000, 3000	4	475.99	1.07	1.12E+03	0.9	850	1.7
Cisapride	3, 10, 30, 100	4	23.54	1.01	11.8	1.3	71	1.8
Ranolazine	3000, 10000, 30000, 100000	4	1.60E+04	0.81	6.52E+03	0.84	3.38E+03	1.1
Verapamil [†]	30, 100, 300, 1000	4	535.74	0.99	502	1.1	172	1.3
Flecainide	100, 300, 1000, 3000	4	871.70	1.05	—	—	—	—

¹: At physiological temperature (Crumb et al. 2016). ²: [Li et al. 2018].
[†]: Each cell was exposed to single drug concentration. NA: Not applicable.

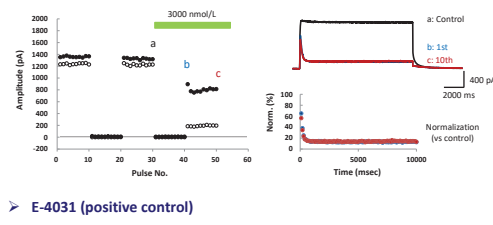
hERG Assay with Dynamic protocol



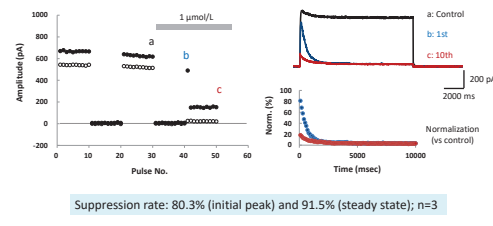
Cisapride



Verapamil



E-4031 (positive control)



Suppression rate: 80.3% (initial peak) and 91.5% (steady state); n=3

Note: An appropriate amount of sucrose was added to the extracellular solution to adjust osmotic balance for the recording using hERG dynamic protocol.

Fractional Block

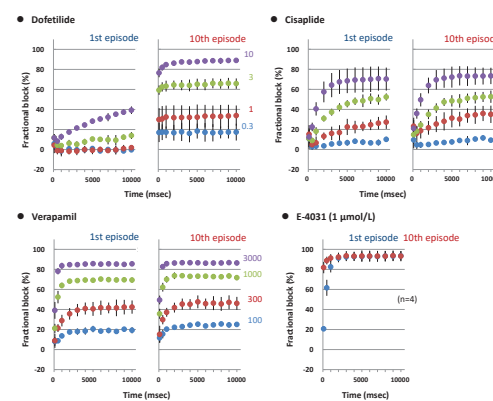
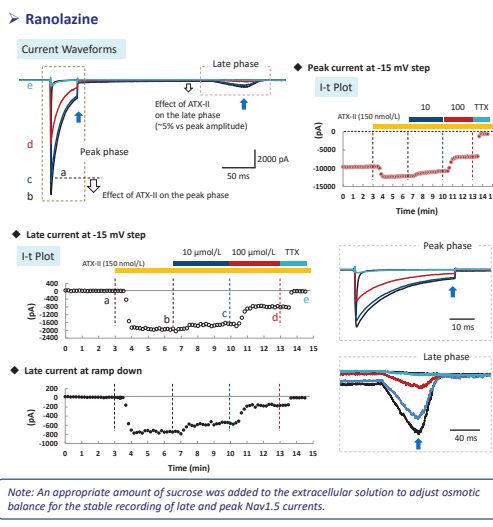


Table 2. Effects of CiPA Compounds on hERG Channel (Dynamic protocol)

TdP risk	Test concentration (μmol/L)	n	Initial peak	Steady state	CiPA data set			
					Manual ¹	Hill	HTS ²	
Bepridil	3, 10, 30, 100	3	68.45	0.46	15.75	0.92	50	0.5
Dofetilide ^a	0.3, 1, 3, 10	3	2.53	0.86	1.85	1.05	4.9	0.9
Astemizole	0.1, 0.3, 1, 3	3	0.77	0.84	0.57	0.95	—	—
Chlorpromazine	100, 300, 1000, 3000	3-4	574.86	0.77	466.40	1.00	929	0.8
Cisapride	3, 10, 30, 100	3-4	NA	NA	8.39	0.80	20	0.7
Ranolazine	1000, 3000, 10000, 30000	3-4	1.35E+04	0.67	8.55E+03	0.89	8.77E+03	0.8
Verapamil	100, 300, 1000, 3000	4	2.48E+03	0.60	369.12	0.85	288	1
Flecainide	300, 1000, 3000, 10000	4	2.47E+03	0.65	1.16E+03	0.86	—	—

¹: [Li et al. 2018]. NA: Not applicable.

Late Nav1.5 Assay Using ATX-II



Note: An appropriate amount of sucrose was added to the extracellular solution to adjust osmotic balance for the stable recording of late and peak Nav1.5 currents.

Table 3. Effects of CiPA Compounds on Late Current of Nav1.5 Channel

TdP risk	Test concentration (μmol/L)	n	IC50	Hill	CiPA data set			
					Manual ¹	Hill	HTS ²	
Bepridil	0.1, 0.3, 1, 3	4	1.32	1.47	0.67	1.31	1.82	1.4
Dofetilide ^a	100	2	>>100 [†]	NA	>>100 [†]	NA	126	1.1
Astemizole ^a	0.1, 0.3, 1, 3	3	0.67	1.22	0.99	1.13	30.3	2.3
Chlorpromazine ^a	0.1, 0.3, 1, 3	4	2.35	1.67	0.76	1.30	4.59	0.94
Cisapride	0.3, 1, 3, 10	3	4.95	1.02	2.55	1.12	9.26E+03	6.3
Ranolazine	3, 10, 30, 100	3-4	82.54	1.31	38.32	1.05	7.94	0.95
Verapamil [†]	1, 3, 10, 30	3	18.99	0.94	8.47	0.95	24.1	2
Flecainide	0.3, 1, 3, 10	3	2.97	1.00	1.70	1.11	—	—

¹: At physiological temperature (Crumb et al. 2016). ²: [Li et al. 2018]. [†]: 0.6% (-15 mV step) and 5.8% (ramp down) at 100 μmol/L.

Peak Nav1.5 Assay

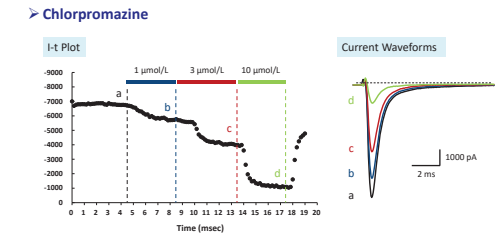
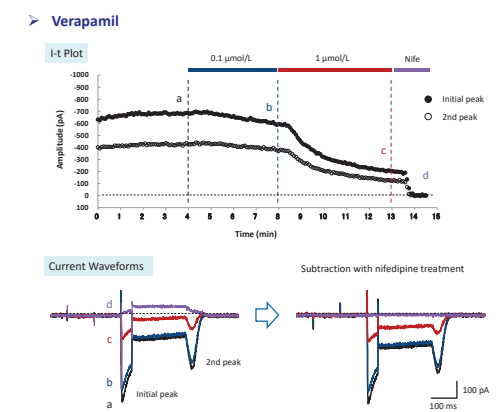


Table 4. Effects of CiPA Compounds on Peak Current of Nav1.5 Channel

TdP risk	Test concentration (μmol/L)	n	IC50	Hill	CiPA data set			
					Manual ¹	Hill	HTS ²	
Bepridil	0.3, 1, 3, 10	4	2.07	1.08	2.96	1.2	1.61E+03	5.4
Dofetilide ^a	100	2	>>100 [†]	NA	>>100 [†]	NA	1.46E+03	5.1
Astemizole ^a	1, 1, 10	4	4.76	1.45	5.41	0.76	7.49	4.8
Chlorpromazine ^a	1, 3, 10, 30	5	3.99	1.54	4.58	2.1	21.2	2.1
Cisapride	1, 10, 50	4	13.05	1.13	1.79E+03	0.67	36.8	2.3
Ranolazine	10, 100, 300, 1000	4-5	239.52	1.23	53.3	1.8	83.7	1.1
Verapamil [†]	10, 30, 100	4	41.43	1.19	2.98E+03	3.5	2.48E+03	5.1
Flecainide	3, 10, 100	4	24.28	1.12	—	—	—	—

¹: At physiological temperature (Crumb et al. 2016). ²: [Li et al. 2018]. [†]: 4.4% at 100 μmol/L.

Cav1.2 Assay



Note: In Cav1.2 current protocol, subtraction with a trace recorded in the presence of nifedipine was conducted for more accurate analyses.

Table 5. Effects of CiPA Compounds on Cav1.2 Channel

TdP risk	Test concentration (μmol/L)	n	Initial peak	2nd peak	CiPA data set			
					Manual ¹	Hill	HTS ²	
Bepridil	0.1, 0.3, 1, 3	4	0.39	1.09	0.36	1.22	2.82	0.65
Dofetilide ^a	100	4	>>100 [†]	NA	>>100 [†]	NA	44.5	3.6
Astemizole ^a	0.05, 0.1, 0.3, 1	4	0.19	1.20	0.22	1.18	0.553	1.2
Chlorpromazine ^a	0.1, 0.3, 1, 3	4-5	0.76	1.59	0.73	1.68	6.30	0.85
Cisapride	0.1, 1, 10, 100	4-5	2.65	1.08	1.49	1.24	0.08E+03	4.8
Ranolazine	30, 100, 300, 1000	4-5	513.65	0.99	324.19	0.92	900	3.9
Verapamil [†]	0.1, 0.3, 1, 3	4	0.39	0.98	0.51	1.02	0.204	1.1
Flecainide	3, 6, 30, 60	4	23.76	0.98	21.46	1.00	—	—

¹: Action potential protocol, Ba²⁺-free charge carrier, physiological temperature (Crumb et al. 2016). ²: [Li et al. 2018].

[†]: 8.6% at 100 μmol/L. ^a: 5.7% at 100 μmol/L.

[†]: Each cell was exposed to single drug concentration. NA: Not applicable.

Conclusions

- In hERG IC50 only and dynamic model protocols, the IC50 values were almost similar to the results by CiPA.
- In late and peak Nav1.5 current protocols, the IC50 values of dofetilide and cisapride deviated far from the results by CiPA.
- Also in Cav1.2 current protocol, the IC50 values of dofetilide and cisapride deviated far from the results by CiPA.
- In silico prediction will be performed with the data set obtained in the research, and the parameters in the analyses will be compared to CiPA work. In addition to CiPA drugs, the TdP risk of flecainide will also be estimated.

References

- Recommended voltage protocols to study drug-cardiac ion channel interactions using recombinant cell lines
- Journal of Pharmacological and Toxicological Methods, Crumb et al., 2016
- Circ Arrhythm Electrophysiol, Li et al., 2017
- CLINICAL PHARMACOLOGY & THERAPEUTICS, Li et al., 2018

Acknowledgement

We would like to express our gratitude to iSmart C-Quest for their generous technical advices and constant discussions with respect to data acquisition and analyses.