Patch-Clamp Studies in the Evaluation of Suppressive Effects of Drugs Fujisawa on the hERG Potassium Channel



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1. Introduction

The Health and Environmental Sciences Institute, which is part of the International Life Sciences Institute (ILSI/HESI), has been building a database of the ion channel assay using a cultured cell line transfected with the human-ether-go-go-related gene (hERG) on standard drugs at two independent facilities. This work was conducted as a S7B activity of the International Conference for Harmonization (ICH) for establishing global guidelines for Safety Pharmacology Study. The data was reviewed at the ILSI Workshop in Washington, DC in June, 2003 and the ICH-Expert Working Group (EWG) Meeting of S7B and E14 in Brussels in July, 2003. Based on the results, it was revealed that there was an approximate 3-fold difference in IC₅₀ values of positive drugs between the two facilities. The inter-facility difference in sensitivity of the hERG assay has become a matter of concern.

This study was conducted by a third party to provide additional information and to encourage further discussions at ICH regarding the inter-facility differences in sensitivity of the hERG assay. The whole-cell patch-clamp technique using HEK293 cells transfected with hERG was used to evaluate the effects of Torsades de Pointes (TdP) positive and negative pharmaceuticals investigated by ILSI/HESI. Since the test procedure employed in this study is slightly different from those used in the evaluation by ILSI/HESI, a side-by-side comparison was conducted between these two procedures in order to determine if there is any influence on the results. Furthermore, other drugs such as E-4031, Sotalol, Quinidine, Lidocaine and Astemizole investigated by QT PRODACT in Japan were evaluated in order to provide reference data for discussion.





Test methods employed to determine the suppressive effects of test substances on the hERG potassium channel transfected in HEK293 cells. A; A schematic diagram of the test substance application system. B; The voltage clamp protocol and currents recorded from a hERG-transfected cell. The cell was held at -70 mV and depolarized to voltages between -50 to 40 mV for 0.5 to 3 sec, and then clamped to -50 mV for 0.75 sec. C; I-V plots of the hERG currents measured at the end of the depolarizing step and the peak tail currents when the voltage protocol described in C was run. D; The voltage clamp protocol and currents recorded from a hERG-transfected cell. The cell was depolarized to 0 mV for 0.75 sec to activate and slightly inactivate the hERG potassium channels, then repolarized to different voltages from -80 to 50 mV for 0.75 sec. E; I-V plots of the peak tail currents when the voltage protocol described in D was run. The temperature was maintained at $37\pm1^{\circ}$ C.







Effects of terfenadine on the hERG current under different test conditions. A; Superimposed current traces in an experiment following the test procedure used in the studies conducted by ILSI/HESI. The pulse command shown in the inset was given to the cell. Currents were elicited by a 1 sec depolarizing step to +20 mV from a holding potential of -80 mV, followed by a repolarizing ramp (-0.5 V/sec) to -80 mV. These pulses were given every 15 sec. Current records from a single HEK293 cell before (open circle) and 10 min after application of terfenadine at 30 nmol/L (closed circle) were superimposed. B; Superimposed current traces in an experiment using the standard test procedure employed at DSTC. The pulse command shown in the inset was given to the cell every 15 sec. Current records from a single HEK293 cell before (open circle) and 10 min after application of terfenadine at 30 nmol/L (closed circle) were superimposed. C; Effects of terfenadine on the concentration-response relationships for the hERG channel in HEK293 cells. The percent suppression of the hERG current under respective testing conditions is plotted as a function of the drug concentration. The IC₅₀ values of terfenadine under ILSI/HESI and DSTC procedures are estimated to be 6 and 18 nmol/L, respectively.

Table 1. Positive controls investigated by ILSI/HESI				Table 2. Negative controls investigated by ILSI/HESI				Table 3. Test substances investigated by QT PRODACT	
IC ₅₀ value (nmol/L)			IC ₅₀ value (nmol/L)			T () (IC ₅₀ value (nmol/L)	
ChanTest *	Zenas *	DSTC	- Test substance -	ChanTest *	Zenas *	DSTC		DSTC	
24	84	17	Amoxicillin	N.A.	N.A.	>100000	E-4031	20	
21	31	26	Aspirin	N.A.	N.A.	>100000	Sotalol	149860	
27	93	25	Captopril	N.A.	N.A.	>100000	Quinidine	511	
1	35	2	Propranolol	8200	2800	6658	Quintume	514	
74	256	43	Diphenhydramine	1900	5700	997	Lidocaine	>100000	
9	33	18	Verapamil	180	252	299	Astemizole	3	
	ontrols investigat IC ₅ ChanTest * 24 21 27 1 74 9	ontrols investigated by ILSI/HES IC ₅₀ value (nmol/L ChanTest * Zenas * 24 84 21 31 27 93 1 35 74 256 9 33	ontrols investigated by ILSI/HESI IC ₅₀ value (nmol/L) ChanTest * Zenas * DSTC 24 84 17 21 31 26 27 93 25 1 35 2 74 256 43 9 33 18	Table 2. Negative ofIC50 value (nmol/L)ChanTest *Zenas *DSTC248417Amoxicillin213126Aspirin279325Captopril1352Propranolol7425643Diphenhydramine93318Verapamil	Table 2. Negative controls investigated by ILSI/HESITable 2. Negative controls investigated by ILSI/HESIIC50 value (nmol/L)Test substanceIC5ChanTest *Zenas *DSTCTest substanceIC5248417AmoxicillinN.A.213126AspirinN.A.279325CaptoprilN.A.1352Propranolol82007425643Diphenhydramine190093318Verapamil180	Table 2. Negative controls investigated by ILSI/HESITable 2. Negative controls investigated by ILSI/HEIC50 value (nmol/L)IC50 value (nmol/L)ChanTest *Zenas *DSTC248417AmoxicillinN.A.N.A.213126AspirinN.A.N.A.279325CaptoprilN.A.N.A.1352Propranolol820028007425643Diphenhydramine1900570093318Verapamil180252	Table 2. Negative controls investigated by ILSI/HESITable 2. Negative controls investigated by ILSI/HESIIC ₅₀ value (nmol/L)ChanTest *Zenas *DSTC248417AmoxicillinN.A.N.A.>100000213126AspirinN.A.N.A.>100000279325CaptoprilN.A.N.A.>1000001352Propranolol8200280066587425643Diphenhydramine1900570099793318Verapamil180252299	Table 2. Negative controls investigated by ILSI/HESITable 3. Test sub by QT PRODACTIC ₅₀ value (nmol/L)Table 2. Negative controls investigated by ILSI/HESITable 3. Test sub by QT PRODACTChanTest *Zenas *DSTCIC ₅₀ value (nmol/L)Test substanceIC ₅₀ value (nmol/L)Test substanceTest substance248417AmoxicillinN.A.N.A.N.A.>100000E-4031213126AspirinN.A.N.A.>100000Sotalol279325CaptoprilN.A.N.A.>100000Quinidine1352Propranolol820028006658Lidocaine7425643Diphenhydramine19005700997Astemizole93318Verapamil180252299Astemizole	

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The IC₅₀ values for suppressive effects of test substances on the hERG channel obtained in this study were all similar to either or both of the two data previously presented by ILSI/HESI. The difference of the test procedures between ILSI/HESI and DSTC for the evaluation resulted in little variance of IC₅₀ values of terfenadine. These results suggest that the inter-facility difference in sensitivity of the hERG assay is limited and the assay would be reliable as a preclinical global standard to determine the QT prolongation potency of drugs.

The authors thank the ILSI/HESI Non Clinical Cardiovascular Subcommittee for their understanding and support of this project. We also highly appreciate the useful comments and encouragement provided by the leadership of QT PRODACT.

6. Summary

7. Conclusion

Acknowledgement